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1	Method Validation for the Analysis of <i>Licorice</i> Acid
2	in the Blending Process by Near Infrared Diffuse
3	Reflectance Spectroscopy
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9	Abstract:
10	The present work described validation of near infrared (NIR) method for the
11	quantification of the concentration of <i>Licorice</i> acid in the blending process of <i>Licorice</i>
12	and talcum powder mixtures. The NIR diffuse reflectance spectra of samples were
13	collected during the mixing process and the partial least square (PLS) model was
14	developed. The accuracy profile (AP) approach that was fully compliant with the ICH
15	Q2 (R1) guideline was used in order to assess the validity of the NIR chemometric
16	method. Particularly, the β -content, γ -confidence tolerance interval, instead of
17	β -expectation tolerance interval in the AP methodology, was introduced to provide a
18	better estimate of measurement risk. The quantitative validation criteria such as
19	trueness, precision (both repeatability and intermediate precision), results accuracy
20	and valid range were obtained. The lower limit of quantification (LLOQ) was 1.26
21	$mg \cdot g^{-1}$. Results demonstrated that NIR spectroscopy is suitable for the analysis of the
22	concentration of <i>Licorice</i> acid. And the risk of using the established analytical method

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in routine phase could be well visualized and controlled. Key words: NIR diffuse reflectance spectroscopy; Method validation; *Licorice* acid; Accuracy profile; β -content, γ -confidence tolerance interval **1. Introduction** Near infrared (NIR) spectroscopy has become a widely used analytical technique in pharmaceutical industry due to its high speed acquisition, non-destructive nature, capacity to measure both physical and chemical properties, and the fact that it needs little or no sample preparation[1]. Therefore, near infrared spectroscopy (NIRS) is more and more considered as an attractive and promising analytical tool for process analytical technology (PAT). Once a calibration model for NIR analysis is developed and favorable predictions are expected, the method must be validated to comply with regulatory requirements. Like any analytical methods, the validation of NIRS method is a mandatory step at the end of the method development in order to give enough guarantees that each of future results during routine use will be close enough to the true value [2]. Generally, validation strategies can be classified into two possible approaches: the traditional approach and the accuracy profile approach. The traditional approach relies on the validation of specific aspects of the method performance step by step, such as accuracy, precision (repeatability and intermediate precision), linearity, range

of application, etc. These validation parameters are consistent with recommendations

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45	of international conference of harmonization (ICH) and other regulatory guidelines
46	from EMA or FDA regulations. Examples of NIRS method validated following this
47	strategy can be found in the determination of water content or API (active
48	pharmaceutical ingredient) content of drug products[3-5], and in the quantification of
49	excipients of antifungal and antibacterial agents[6, 7]. But this type of validation is
50	time consuming and laborious. Furthermore, this strategy can be concluded wrongly
51	that a method giving imprecise results can be more easily validated than a precise one
52	[8].

The accuracy profile (AP) validation protocol was brought forward by the SFSTP (La Societé Francaise des Sciences et Techniques Pharmaceutiques) [9-11]. Accuracy profile is based on the combination in the same graph of the tolerance interval and the acceptable limits, and circumvents some drawbacks of the traditional validation procedures. Compared with the traditional approach, the accuracy profile approach not only simplifies the validation process of an analytical procedure, but also allows monitoring the risk of utilization. Moreover, it can declared an analytical method is valid or not, and the analytical result is guaranteed to be fit for the intended purpose of the analytical method [12, 13]. Several applications of NIR spectroscopy used this approach can be found as follows. Schaefer used the accuracy profile approach to validate the on-line NIR method to control an API crystallization step [14]. Tomuta used this approach to demonstrate that the NIR chemometric methods meet the requirements of a high throughput method for the determination of drug content and pharmaceutical properties of indapamide tablets [15]. Wu successfully

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used PLS model and accuracy profile method for accurate determination of chlorogenic acid content in L. japonica solution in the ethanol precipitation process [16]. Ziémons studied to develop a robust NIR calibration model to determine the acetaminophen content of a low-dose syrup formulation, where the accuracy profile confirmed the adequate accuracy of results generated by the method all over the investigated API concentration range [17]. Fonteyne assessed the in-line moisture content during the drying process in a six-segmented fluid bed dryer of a continuous tablet production line by the accuracy profile, and it was statistically demonstrated that the new NIR method performed at least as good as the Karl Fischer reference method [18]. The AP validation strategy is also fully compliant with the ICH Q2 (R1) guideline [12], and the prediction interval is built by the β -expectation tolerance interval.

However, Saffaj recently reported that the β -expectation tolerance interval cannot accurately predicted future measurements of the method in routine phase since it was incapable to assess the routine uncertainty rightly and was unfortunately not able to protect the laboratory and the client interests at the same time [20-22]. Since the β -expectation tolerance interval only contains the information of trueness and precision about the analysis method, it may underestimate the measurement risk. While, the β -content, γ -confidence tolerance interval provides a good estimate of measurement risk, and gives the best guarantees concerning the decision of declaring a method as valid.

Therefore, in the present work, the β -content, γ -confidence tolerance interval is

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recommended to build the accuracy profile for the validation of NIR analytical method. And the NIR quantitative analysis of the concentration of *Licorice* acid in the mixture of *Licorice* and talcum powder collected during the blending process was taken as the research object. The aim of this study is to apply the new validation strategy to study whether the NIR spectroscopy is suitable for the analysis of the concentration of *Licorice* acid.

2. Theory

Accuracy profile is a graphical decision making tool aiming to help the analyst in deciding whether an analytical procedure is valid. It is 2D-graphical representation results for trueness, tolerance intervals and acceptance limits [8]. Whereas, validation must cover up the whole application domain of the method. The trueness and precision are needed to be calculated at each concentration levels. This ideal acceptance criteria would ensure that a high proportion (say β) of future observations lie within acceptance limits (say λ), with a high degree of confidence (say γ), where the β -content, γ -confidence tolerance interval could help fulfill this task [29].

2.1 Estimation of trueness and precision

The $I \times J \times K$ full factorial validation protocol was utilized to design the validation data set, where the effect of three aspects, i.e. conditions (*I*), the number of repetitions (*J*) and level of concentrations (*K*) were taken into account [23]. The estimate of the trueness and precision of the method was carried out at each of the considered *k* concentration levels, using the following statistical model [11]:

- 5 -

111
$$Y_{ij} = \mu + \alpha_i + \varepsilon_{ij} (i = 1, 2, ..., m; j = 1, 2, ..., n)$$
(1)

112 Where, Y_{ij} is the *j*-th measured value of the *i*-th condition at the concentration 113 level *k*. μ is the mean of the measured values at each concentration level. *m* is the 114 number of series, *n* is the number of independent replicates per series. α_i is the 115 difference between the *i*-th series average and the μ at level *k*. α_i is considered as a 116 normal random variable with 0 as the average and $\hat{\sigma}_B^2$ as the variance. ε_{ij} is the 117 experimental error considered as a normal random variable with an average of 0 and a 118 variance of $\hat{\sigma}_E^2$.

The experimental error is supposed to be independent of the series. The $\hat{\sigma}_B^2$ and $\hat{\sigma}_E^2$ variances represent the inter-series and intra-series variances, respectively. The restricted maximum likelihood method is used to estimate, at every concentration level, the parameters u_k , $\hat{\sigma}_B^2$ and $\hat{\sigma}_E^2$ of the model. Define MS_B and MS_E the mean square of inter-series and intra-series, respectively.

124
$$MS_B = \frac{n}{m-1} \sum_{i=1}^{m} (\bar{Y}_i - \bar{Y})^2$$
(2)

125 Where $\overline{Y}_{i} = \frac{1}{n} \sum_{j=1}^{n} Y_{ij}$, $\overline{Y} = \frac{1}{mn} \sum_{i=1}^{m} \sum_{j=1}^{n} Y_{ij}$

126
$$MS_E = \frac{1}{m(n-1)} \sum_{i=1}^m \sum_{j=1}^n (Y_{ij} - \overline{Y}_i)^2$$
(3)

127 If
$$MS_E < MS_B$$
, then

128
$$\hat{\sigma}_B^2 = \frac{MS_B - MS_E}{n} \tag{4}$$

$$\hat{\sigma}_E^2 = MS_E \tag{5}$$

130 Otherwise

$$\hat{\sigma}_B^2 = 0 \tag{6}$$

132
$$\hat{\sigma}_E^2 = \frac{1}{mn-1} \sum_{i=1}^m \sum_{j=1}^n (Y_{ij} - \overline{Y})^2$$
(7)

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133 Precision:

The ICH defines precision as the closeness of agreement between a series of measurements obtained from multiple sampling of the same homogeneous sample under the prescribed condition. Precision is evaluated at two levels: repeatability and intermediate precision [19].

Repeatability expresses the precision under the same operating conditions over a short interval of time, and should be assessed using a minimum of 9 determinations covering the specified range for the procedure [12]. Therefore, the intra-series variance in Eq. (5) or Eq. (7) provides the repeatability variance estimate:

142 Repeatability:
$$\hat{\sigma}_{Re}^2 = \hat{\sigma}_E^2$$
 (8)

143 Intermediate precision expresses within laboratories variations generated from 144 different equipment, different days or different analysts. The sum of intra- and 145 inter-series variance provides an estimation of the intermediate precision:

146 Intermediate precision:
$$\hat{\sigma}_M^2 = \hat{\sigma}_B^2 + \hat{\sigma}_E^2$$
 (9)

147 **Trueness:**

The trueness of an analytical procedure, also called theoretical true value, express the closeness of agreement between the average of the results calculated by the method and the accepted reference value [19]. The trueness is expressed as bias and recovery in relative form.

152
$$bias(\%) = \frac{\overline{Y} - Xr}{Xr} \times 100$$
(10)

153
$$re \operatorname{cov} ery(\%) = \frac{\overline{Y}}{Xr} \times 100$$
 (11)

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1

154 Where, Xr is the theoretical value.

155 **2.2 Estimation of the** β **-content,** γ **-confidence tolerance interval**

156 The β -content, γ -confidence tolerance interval is defined as follows [24]:

157
$$P(P[L \le x_i \le U] \ge \beta) = \gamma$$
(12)

158 The β -content, γ -confidence tolerance interval provides lower (*L*) and upper (*U*) 159 limits that claim a specified proportion β of assayed values will lie within the interval 160 [L, U], with specified confidence level γ . For example, for $\beta = 0.70$ and $\gamma = 0.95$, the 161 β -content, γ -confidence tolerance interval expresses that there is a probability (*P*) of 162 0.95 that 70% of the individual observations of the population are included in the 163 interval [*L*, *U*].

164 According to the Mee's approach [25], the β -content, γ -confidence tolerance 165 interval under this method takes the following form:

166
$$[L,U] = \left[\overline{Y} - k_C \hat{\sigma}_M; \overline{Y} + k_C \hat{\sigma}_M\right]$$
(13)

167 With

168
$$k_{C} = \sqrt{\frac{\nu' \chi_{1;\beta}^{2}(\tau)}{\chi_{\nu';1-\gamma}^{2}}}$$
(14)

In Eq. (13), k_c represents a Chi-square distribution associated with β and γ for interval estimation [25]. $\chi^2_{1;\beta}(\tau)$ is the β quantile of a noncentral Chi-square distribution with the degree of freedom 1. τ is noncentrality parameter. $\chi^2_{\nu;1-\gamma}$ denotes the 1- γ quantile of a non-central Chi-square distribution with degrees of freedom ν' .

174 And

accuracy.

175
$$v' = \frac{(R'+1)^2}{(R'+(1/n))^2/(m-1)+(1-(1/n))/mn}$$
(15)

176
$$\tau = \frac{1}{mnB'}$$
 (16)

177
$$R' = \max\left[0, \frac{1}{n}(\frac{F}{F_{\eta}} - 1)\right]$$
(17)

178 And
$$B' = \frac{R'+1}{nR'+1}$$
 (18)

Where *F* is the mean square ratio MSb/MSe, and *F_η* is the 100η percentile of an *F* distribution with v₁ = m (n-1) and v₂ = (m-1). However, based on numerical results,
the recommended values of η are 0.85, 0.905 and 0.975, corresponding to γ = 0.90,
0.95 and 0.99, respectively [26].
Thus, the β-content, γ-confidence tolerance interval can be rewritten in relative

184 form as follows:

$$[L(\%), U(\%)] = [bias(\%) - k_C RSD(\%), bias(\%) + k_C RSD(\%)]$$
(19)

186 Where:

187
$$RSD(\%) = \frac{\hat{\sigma}_M}{X_r} \times 100 \tag{20}$$

2.3 Establishment of the accuracy profile

189 The proposed building procedures of accuracy profile are as follows:

190 1) Set acceptance limits
$$(-\lambda, +\lambda)$$
.

191 2) Construct β -content, γ -confidence tolerance intervals ([L, U] or [L (%), U (%)])

192 for each level according to Eq. (13) or Eq. (19) with desired confidence level γ

193 3) Make a 2D-graphical representation of results with the horizontal axis for the 194 concentration levels and vertical axis for the tolerance interval limits (L, U) and

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196	4) Compare the tolerance interval limits (<i>L</i> , <i>U</i>) to the acceptance limits (- λ , + λ)
197	5) If (L, U) falls completely within $(-\lambda, +\lambda)$, the method is accepted; otherwise,
198	the method is not accepted
199	
200	3. Experimental
201	3.1 Materials
202	Licorice powder (lot number: 20120926) and medicinal talc (lot number:
203	20120514) were purchased from Ben Cao Fang Yuan Medicine Co., Ltd. (Beijing,
204	China). Licorice acid monoammonium salt (lot number: 111229) was supplied by
205	National Institutes for Food and Drug Control (Beijing, China). HPLC grade
206	methanol and phosphoric acid were purchased from Fisher Scientific (USA). HPLC
207	grade ammonium dihydrogen phosphate was purchased from Acros Organics (USA),
208	and the pure water was purchased from Wahaha Co., Ltd. (Hangzhou, China).
209	3.2 Acquisition of spectroscopic data
210	An Antaris near infrared spectrometer (Thermo Fisher Scientific Inc., USA) was
211	used to collect the spectroscopic data. Each spectrum was an average of 64 scans with
212	the resolution 8 cm ⁻¹ over the range $10000 \sim 4000$ cm ⁻¹ . A background spectrum was
213	taken daily in air. And the integrating sphere diffuse mode with rotating sample cup
214	was applied.
215	3.3 Reference method

The reference method used for the *Licorice* acid determination was HPLC assay
recommended by the Chinese Pharmacopoeia (Ch. P., 2010 Edition). An Agilent 1100
HPLC apparatus, equipped with a quaternary solvent delivery system, an auto sampler, -10-

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a DAD detector and HP workstation for data processing were used. The concentration of *Licorice* acid was analyzed by the reverse phase chromatography on an Agilent C18 column (4.6×250 mm, 5µm) with isocratic elution of the mobile phase consisted of methanol, ammonium dihydrogen phosphate buffer (65:35, v/v) at the flow rate of 1.0 mL·min⁻¹. A column temperature of 30 °C, injection volume of 20 µL and detection wavelength at 250 nm were used.

225

3.4 Calibration and validation protocols

The *licorice* and talcum powders with the mass ratio of 1:6 were mixed by the 226 10L three dimensional blender (ZNW-10, Beijing Xing Shi Li He Co., Ltd., China). 227 The filling coefficient was set at 70%, and the spindle speed was 13 rpm. During the 228 mixing process, the blender stopped at 5, 7, 9, 11, 12, 13, 14, 15, 17, 19 mins, and the 229 230 time required to reach a homogeneous blend is 17mins as assured by HPLC analysis. And then, 5 g powders are respectively sampled at 5 positions preset, as shown in 231 Fig.1. These samples were directly analyzed by NIR under the conditions specified in 232 Section 3.2. Two batches of mixing experiments were carried out, and 100 ($10 \times 5 \times 2$) 233 samples were finally got. 234

235

Fig.1

The validation protocol used the $3 \times 5 \times 3$ full factorial experiment design. Five different *Licorice* acid concentrations levels (0.78 mg·g⁻¹, 1.56 mg·g⁻¹, 2.34 mg·g⁻¹, 3.12 mg·g⁻¹ and 3.89 mg·g⁻¹) were investigated, and each level was performed in 3 replicates on 3 different days, resulting in 45 samples in the validation set.

240 3.5 Data Processing

SIMCA-P 11.5 (Umetrics, US) and Unscrambler 7.0 (CAMO, Norway) were used to perform spectral pretreatments. The PLS regression was performed on Matlab version 7.0 (Math Works Inc., USA) with PLS Toolbox 2.1 (Eigenvector Research Inc., USA). The calculation of the β -content, γ -confidence tolerance interval and the construction of the accuracy profile were realized using homemade programs.

- 247 4. Results and discussion
- 248 4.1 NIR method development

Before model building, the first step of NIR method development is outlier detection to improve the performance of the model. First, the raw NIR spectra of 100 samples were analyzed by the principal component analysis (PCA) with the first principal components explaining 87.21% variation and the first two principal components explaining the 99.8% variation of samples. Then, score plots with two principal components are used to identify spectra clusters and to reveal the spatial distribution of samples as shown in Fig. 2A. The Hotelling T^2 ellipse with 95% confidence is calculated to identify the potential outliers as shown in Fig. 2B. As a result, 4 abnormal samples were removed. And the rest 96 samples were divided into the calibration set (56 samples) and the validation set (40 samples) by the Kennard-Stone (K-S) method.

Fig.2

The second step for NIR method development is spectral pretreatment. In order to improve the prediction ability of model, different spectral pretreatment methods

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263	were investigated. Multiplicative signal correction (MSC) and standard normal variate
264	(SNV) were used to eliminate the impact of light scattering generated by the uneven
265	distribution of the particles size. The first derivative (1std) and second derivative
266	(2ndd) treatments for spectral data were used to eliminate the spectral baseline drift,
267	strengthen band characteristics and overcome overlapping bands. The Savitzky-Golay
268	(S-G) smoothing and wavelet de-nosing of spectra (WDS) were used to effectively
269	smooth the high frequency noise, improve the signal to noise ratio and reduce the
270	noise impact. For S-G smoothing, the filter width was set at 9 wavenumbers and the
271	polynomial order was 2. And then, the optimal number of latent variable was
272	optimized using the leave-one-out (LOO) cross validation method. Conventional
273	correlation coefficient $r$ for both calibration and validation sets, the root mean squared
274	error of calibration (RMSEC), the root mean squared error of cross-validation
275	(RMSECV), the root mean squared error of prediction (RMSEP) and ratio of
276	performance deviation (RPD) were used to select several model candidates.
277	Table 1 PLS model characteristics with and without preprocessed spectra.

Drotrootmont	I Va	Calibration set				Validation set			
rieueatment	LVS	$r_{cal}$	RMSEC	RMSECV	BIAS _{cal}	$r_{val}$	RMSEP	RPD	$\operatorname{BIAS}_{\operatorname{val}}$
Origin	9	0.9932	0.085	0.134	0.068	0.9509	0.138	3.08	0.108
S-G	9	0.9921	0.092	0.136	0.072	0.9524	0.135	3.13	0.106
1 std	5	0.9956	0.068	0.162	0.056	0.9473	0.140	3.03	0.105
2ndd	3	0.9757	0.160	0.187	0.118	0.9199	0.173	2.45	0.139
S-G+1std	8	0.9986	0.039	0.166	0.032	0.9495	0.139	3.06	0.099
1std+S-G	5	0.9959	0.066	0.160	0.053	0.9492	0.137	3.10	0.101
MSC	9	0.9925	0.089	0.180	0.073	0.9464	0.144	2.95	0.114
SNV	9	0.9921	0.092	0.179	0.076	0.9462	0.144	2.95	0.115
WDS	10	0.9871	0.117	0.153	0.088	0.9431	0.141	3.00	0.108

278 Note: Origin means using the original spectra

279	As presented in Table 1, after spectral preprocessing, the prediction accuracy of
280	the model is not significantly improved. Compared to other preprocessing methods,
281	the 1std+S-G preprocessing method used relatively fewer PLS factors with values of
282	$r_{cal}$ (0.9959) close to 1, and the smaller values of RMSEC (0.066 mg·g ⁻¹ ), RMSECV
283	$(0.160 \text{mg} \cdot \text{g}^{-1})$ were also indications of the good quantitative performances of the
284	NIRS method developed. The RPD value 3.10 was greater than 3, demonstrating that
285	the predictive performance of the developed NIR calibration model was good.
286	Additionally, variables selection was done after preprocessing of spectra, but the
287	model performance did not improve (the data were shown in Table S1 and the results
288	were shown in Table S2). Therefore, the 1std+S-G preprocessing method was chosen
289	to build the PLS regression model. Fig.3A showed that the model performance
290	changes with the latent variable (LV) factors. And it could be seen that the RMSEC
291	and RMSEP values did not change at 5 LVs, based on which the PLS model was
292	established. The relationship between the calibration set and the prediction set of the
293	regression model was shown in Fig. 3B.
294	Fig. 3

**4.2 NIR method validation** 

In agreement with the guideline of ICH Q2, the typical validation characteristics for assay procedures like accuracy, precision, range, and linearity were determined, and the accuracy profile was established. Due to the quality control of traditional Chinese medicine is similar to that of biological products, the acceptance limits (- $\lambda$ , + $\lambda$ ) were set at ± 20% for the validation of the NIR method [11, 19, 20, 27]. To Page 15 of 27

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compute the  $\beta$ -content,  $\gamma$ -confidence tolerance intervals and build the accuracy profile, he present work had opted for the 4-6- $\lambda$  rule adopted by the FDA for the validation of ioanalytical procedures. And this rule was translated into  $\beta = 66.7\%$  and  $\gamma = 90\%$  by Hoffman and Kringle [28-30].

#### .2.1 Accuracy

Accuracy takes into account the total error which is the sum of systematic and andom errors, related to the validation result. As presented in **Table 2 and Fig. 4**, the ower and upper  $\beta$ -content,  $\gamma$ -confidence tolerances for 1.56 mg·g⁻¹ (-1.82, 8.37), 2.34  $mg \cdot g^{-1}$  (-12.2, 17.1), 3.12  $mg \cdot g^{-1}$  (-6.20, 2.29), 3.89  $mg \cdot g^{-1}$  (-19.4, 4.00) concentration evels were all within the acceptance limits of  $\pm 20\%$ . Consequently, the method can be considered as valid over the concentration range from 1.56 mg  $g^{-1}$  to 3.89 mg  $g^{-1}$ . Nevertheless, the accuracy was outside the acceptance limits for the level 0.78 mg  $g^{-1}$ . This result can be explained by: with a reduced concentration levels, system and andom errors would increase.

#### 4.2.2 Precision

Precision is evaluated at two levels: repeatability and intermediate precision at five concentration levels. The variance of repeatability and time dependent intermediate precision as well as the relative standard deviation (RSD) were calculated from estimated concentrations. From the precision results in Table 2, it is obvious that the intermediate precision is worse than the repeatability, which means that there is an important operator and/or day effect at these concentration levels. As 321 can be seen from Fig. 4, the dispersion of the results is good for 1.56 mg  $g^{-1}$ , 2.34 322

mg·g⁻¹, 3.12 mg·g⁻¹ and 3.89 mg·g⁻¹ concentration levels, leading to good repeatability and intermediate precision values. However, with the decrease of concentration, the repeatability and intermediate precision values significantly increased. The values at the 0.78 mg·g⁻¹ concentration level were too large to satisfy the analytical requirements.

#### **4.2.3 Range**

The intersection between the accuracy profile and the acceptance limits defines the lower limit of quantification (LLOO) as well as the upper limit of quantification (ULOQ) of the procedure. The lower and upper limits of quantification (LLOQ and ULOQ) define the range where an analytical method is able to quantify accurately. They are respectively the smallest and highest concentration levels where the  $\beta$ -content,  $\gamma$ -confidence tolerance intervals are included within the acceptance limits. If the  $\beta$ -content,  $\gamma$ -confidence tolerance intervals never cross the acceptance limits, then the LLOQ and ULOQ are located at the beginning and at the end of the active content range investigated.

In our case, the LLOQ value was  $1.26 \text{ mg} \cdot \text{g}^{-1}$  via interpolation from the accuracy profile (**Fig. 4**), and ULOQ value was  $3.89 \text{ mg} \cdot \text{g}^{-1}$ . So the quantitative range was defined from  $1.26 \text{ mg} \cdot \text{g}^{-1}$  to  $3.89 \text{ mg} \cdot \text{g}^{-1}$ .

Table 2 ICH Q2 (R1) validation criteria for the NIR method

	Moon	Trueness		Precisi	ion	Accuracy	
Level $(mg \cdot g^{-1})$	calculated concentration	Relative bias (%)	Recovery (%)	Repeatability (%)	Intermediate precision (%)	β-CTI (%)	Abs β-CTI
0.78	0.65	-16.8	83.2	9.82	11.7	[-48.9,15.3]	[0.40,0.90]
1.56	1.61	3.28	103.3	2.87	2.87	[-1.82,8.37]	[1.53,1.69]

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	2.34	2.40	2.47	102.5	2.45	4.53	[-12.2,17.1]	[2.05,2.74]
	3.12	3.06	-1.96	98.0	1.40	1.60	[-6.20,2.29]	[2.93,3.19]
	3.89	3.59	-7.68	92.3	1.51	3.50	[-19.4,4.00]	[3.14,4.05]

342 Note: The  $\beta$ -CTI (%) is relative  $\beta$ -content,  $\gamma$ -confidence tolerance interval; Abs  $\beta$ -CTI is absolute

 $\beta$ -content,  $\gamma$ -confidence tolerance interval.

#### Fig.4

#### 345 4.2.4 Linearity

The linearity of an analytical method is its ability within a definite range to obtain results directly proportional to the concentrations (quantities) of the analyte in the sample. Therefore, a linear model was fitted on the calculated concentrations of the validation standards for all series as a function of the introduced concentrations.

The relationship between the NIR predictions and the theoretical values was evaluated by the linear equation:  $v = 0.9426 \times x + 0.0572$  with  $R^2$  of 0.9820. The intercept, the slope and the  $R^2$  values demonstrated good agreement between the NIR predictions and the theoretical values. In order to prove the method linearity, the absolute  $\beta$ -content,  $\gamma$ -confidence tolerance intervals were applied. The linearity over the *Licorice* acid content range  $1.56 \sim 3.89 \text{ mg} \cdot \text{g}^{-1}$  was demonstrated since the  $\beta$ -content,  $\gamma$ -confidence tolerance interval ( $\gamma = 90\%$ ) limits were within the absolute acceptance limits as shown in Fig. 5. 

#### Fig.5

#### **5. Conclusion**

In this paper, a new strategy based on the accuracy profile methodology which incorporates the  $\beta$ -content,  $\gamma$ -confidence tolerance interval has been successfully managed to validate the NIR quantitative analytical procedures in a traditional -17-

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363	Chinese medicine blending process. Results demonstrated the developed NIR method
364	was suitable for the analysis of the concentration of Licorice acid. The proposed
365	approach offered a formal statistical framework by which the performance of the
366	methodwas assessed. The method validation characteristics such as accuracy,
367	precision, range, linearity and limit of quantification could be obtained for customers.
368	In addition, the improved accuracy profile approach gave a good estimate of
369	measurement risk, and provided visual and reliable method decision tool in the
370	validation stage and controlled the risk of using the analytical method in routine phase.
371	Moreover, it is believed that the improved accuracy profile approach is not only
372	suitable for NIR method, but can also be used for other analytic procedures.
272	

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6

ULOD

### **Graphical Abstract**









¹ Theoretical Concentration (³mg.g⁻¹)

-40

LLOD

Range

#### **Figure captions**

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**Figure 2** A Scores plot; **B** Outliers detection by the Hotelling  $T^2$  ellipse.

**Figure3 A** Calibration characteristics vs. number of latent factors; **B** Correlation graph of NIR predictive values with reference values.

**Figure 4** Accuracy profile for the *Licorice* acid content. The red line is the relative bias; the medium dashed lines are the  $\beta$ -content;  $\gamma$ -confidence tolerance intervals ( $\gamma = 90\%$ ) and the red short dashed lines are the acceptance limits (±20%), the 9 black points at each concentration level are relative bias for each predictive value.

Figure 5 Linear profile for NIR analysis of the *Licorice* acid content. The blue medium dashed lines are absolute  $\beta$ -content,  $\gamma$ -confidence tolerance intervals ( $\gamma = 90\%$ ), and red short dashed lines represent the accepted limits at ± 20%. The continuous line is the identity line y = x.

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Fig.4

